Testing the safety-net role of hedgerow tree roots by ¹⁵N placement at different soil depths

E. C. ROWE¹, K. HAIRIAH², K. E. GILLER¹, M. VAN NOORDWIJK³ and G. CADISCH^{1, *}

¹ Department of Biological Sciences, Wye College, University of London, Wye, Ashford, TN25 5AH, UK; ² Fakultas Pertanian, Jurusan Tanah, Universitas Brawijaya, Jalan Veteran, Malang, Indonesia; ³ ICRAF – SE ASIA, PO Box 161, Bogor 16001, Indonesia (*Author for correspondence: E-mail: g.cadisch@wye.ac.uk)

Key words: competition, leaching, N uptake, subsoil, N₂ fixation, Indonesia

Abstract. Trees which root below crops may have a beneficial role in simultaneous agroforestry systems by intercepting and recycling nutrients which leach below the crop rooting zone. They may also compete less strongly for nutrients than trees which root mainly within the same zone as crops. To test these hypotheses we placed highly enriched ¹⁵N-labelled ammonium sulphate at three depths in the soil between mixed hedgerows of the shallow-rooting Gliricidia sepium and the deep rooting Peltophorum dasyrrhachis. A year after the isotope application most of the residual ¹⁵N in the soil remained close to the injection points due to the joint application with a carbon source which promoted ¹⁵N immobilization. Temporal ¹⁵N uptake patterns (twoweekly leaf sub-sampling) as well as total ¹⁵N recovery measurements suggested that Peltophorum obtained more N from the subsoil than Gliricidia. Despite this Gliricidia appeared to compete weakly with the crop for N as it recovered little ¹⁵N from any depth but obtained an estimated 44-58% of its N from atmospheric N2-fixation. Gliricidia took up an estimated 21 kg N ha⁻¹ and Peltophorum an estimated 42 kg N ha⁻¹ from beneath the main crop rooting zone. The results demonstrate that direct placement of ¹⁵N can be used to identify N sourcing by trees and crops in simultaneous agroforestry systems, although the heterogeneity of tree root distributions needs to be taken into account when designing experiments.

Introduction

Where crop roots are restricted, as at early crop growth stages or in soils which have chemical or physical barriers to root growth at depth, nutrients are easily leached beyond the reach of crop roots. This is particularly true of mineral nitrogen, which is poorly retained in soils. The presence of trees in fields will improve overall nutrient use efficiency if these trees actively take up nutrients which would otherwise have been lost by leaching (Young, 1997). The idea that recycling by trees of nutrients which have leached beyond the range of crop roots is of benefit to the nutrient balance of a field has become known as the safety – net hypothesis (van Noordwijk et al., 1996; van Noordwijk et al., 1992). This may be contrasted with a 'nutrient pump' role, where trees are taking up nutrients from deep soil layers or groundwater which have not recently passed through the crop rooting zone (Cannell et al., 1996).

N dynamics in mixed cropping systems are complex. Trees may compete

with crops for N in the surface soil layers. Spatial and temporal patterns of N release and uptake will be affected by such factors as the timing of N additions, the quality of pruning materials, and the degree of synchrony of crop and tree demand (Haudayanto et al., 1997; Xu et al., 1993). In assessing the overall effect of incorporation of trees into cropping systems, the benefit of improved N use efficiency must be set against competition for and uptake of N which would otherwise have been used by the crop (Cannell et al., 1996). In general, good intercropping trees are likely to be those whose spatiotemporal pattern of N uptake overlaps little with that of the crop.

Prediction of yield from a mixed cropping system also requires estimation of other interactions, such as shading effects, and competition for water. The design of productive, locally adapted agroforestry systems will remain largely a process of trial and error unless a mechanistic understanding of these processes is reached. The development of complex models is helping to facilitate understanding of the overall effects of tree – crop interactions, but must be accompanied by field validation (Cadisch et al., 1997; van Noordwijk, 1996). This study was designed in part to provide data suitable for comparison with simulated data produced using the WaNuLCAS model (van Noordwijk and Lusiana, 1997). This model incorporates a detailed treatment of soil N processes and plant uptake from up to 12 soil compartments.

Hedgerow intercropping trials in North Lampung showed that *Peltophorum* dasyrrhachis, a relatively slow growing, non-nodulating leguminous tree, had a stronger positive effect on sustained crop yields than *Gliricidia sepium*, a tree which fixes nitrogen and produces abundant high quality prunings (van Noordwijk et al., 1997a). *Peltophorum* has a sparse root system which extends deeply into the soil. In contrast, *Gliricidia* has a dense, shallow root system largely coincident with that of annual crop species (Akiefnawati 1995; Hairiah et al., 1992). The present study was designed to test the specific hypotheses that the more deeply rooted *Peltophorum* takes up more N from the subsoil, and competes with the crop less strongly for topsoil N, than *Gliricidia*.

A direct test of root activity was made, by placement of ¹⁵N at different soil depths in the hedgerow intercropping system and measurement of relative recovery by trees and crops. In studies of root activity distributions, ³²P has been more generally used than ¹⁵N (IAEA, 1975). Phosphorus is likely to remain within a relatively restricted soil location, in contrast to N which is highly mobile in soils. Both ionic forms of N have low adsorption coefficients (defined as the ratio of adsorbed ions to ions in solution) and are therefore subject to rapid leaching (Wong et al., 1990). The mobility of N may be limited by the addition of a carbon source to stimulate immobilization and gradual re-release of labelled nitrogen (Witty and Ritz, 1984), and this was the method used here.

Materials and me hods

Study site

The experiment was set up on a farmer's field near the field station of Universitas Brawijaya/ICRAF in North Lampung. Sumatera, Indonesia $(4^{\circ}31' \text{ S}, 104^{\circ}55' \text{ E})$. Rainfall is 2000–2500 mm annually, with a wet season extending from November to May. Temperatures fluctuate little throughout the year, generally remaining in the range 25–35 °C (Van der Heide et al., 1992). The experiment was carried out during the second cropping cycle of the wet season.

The soils in the study area are Typic Kandiudults with pH (H₂O) around 5.2 in the topsoil and 4.8 in the subsoil. Aluminium in the subsoil solution reaches 106 μ mol l⁻¹ (Van der Heide et al., 1992). Total N varies from around 0.15% in the topsoil to 0.05% in the subsoil. Low pH and high concentrations of aluminium in the subsoil, associated with the presence of a plinthitic layer, severely restrict rooting depth of many species (Hairiah et al., 1991).

Hedgerows of *Gliricidia* and *Peltophorum* were established in the field four years previously. *Gliricidia* was established by planting stakes, *Peltophorum* by transplanting wildings (spontaneous seedlings). Alternating hedgerows of *Peltophorum* and *Gliricidia* were spaced 4 m apart, with a distance of 0.5 m between trees in the hedgerow. Van Noordwijk et al. (1997b) gave details of the establishment phase of this trial. Groundnut was sown on 15th April 1995. Seven rows were sown per alley, at a spacing of 0.5×0.25 m. Hedgerows were pruned before sowing, either at ground level or at 0.75 m stem height, and the prunings spread on the crop alley.

¹⁵N recovery experiment

Placements of ¹⁵N were made in the centre of the crop alley (between alternating *Gliricidia* and *Peltophorum* hedgerows), on 12th May 1995, 27 days after sowing the groundnut. Three placements were made at 5 cm and 35 cm soil depths, and two at 55 cm depth. Placements at the same soil depth were separated by a minimum of 4 m, and were made within the same alley. The separation between placements at different depths was at least 30 m. This compromise on the randomness of the design was considered essential to reduce the risk of contamination between treatments, particularly the chance that large recoveries would be attributed to deep placements because of horizontal foraging from shallow placements. Limitations of space, and the need for sufficient separation between placements, restricted the number of replications possible. For this reason the effects of hedgerow pruning height have been disregarded in this study.

Ammonium sulphate (20 atom %¹⁵N) was applied to the soil through plastic tubes installed in holes pre-augered to the required depth, ensuring a tight fit with the surrounding soil. Five tubes were installed per application site,

25 cm apart, in a 1 m strip along the centre of the crop alley. A 25 ml solution of 0.4 g N, mixed with sucrose to give a C:N ratio of 8:1, was applied through each tube. The carbon source was intended to stimulate microbial immobilization of the ¹⁵N, maintaining it within a limited soil volume. Total N applied per site was thus 2 g, equivalent to approximately 20 kg ha⁻¹. Plastic tubes were capped after application.

Sample collection

Leaf samples were taken from trees and groundnut plants before ¹⁵N placement and approximately every two weeks thereafter. Samples were taken from the first fully formed leaves on each shoot.

In each replicate, trees and crop plants from zones at different distances from ¹⁵N placements were sampled separately. Hedgerows on each side of a placement were either of *Gliricidia* or *Peltophorum*. Hedgerow trees were sampled in two zones; the three trees perpendicular to a placement site ('centre') and three trees either side of these ('border': six trees). The six crop plants in the centre row adjacent to placement tubes were sampled ('centre'), six plants in rows on either side of these ('border': 12 plants), and two plants at either end of each of the 'centre' and 'border' rows (Zone 3: 12 plants) (Figure 1).



¹⁵N injection tube

Figure 1. Layout of ¹⁵N application plots in a hedgerow intercropping system at Brawijaya University in North Lampung, Sumatera, Indonesia.

84

Soil from the ¹⁵N application points was sampled by augering 406 days after application. Samples were taken from beneath the five application tubes, and mixed to make composite samples for each depth interval. Samples were air dried and analysed for ¹⁵N.

The ¹⁵N enrichment of plant and soil samples was determined using a Europa 20/20 mass spectrometer (Europa Scientific, Crewe, UK) coupled to an automated C/N analyzer. ¹⁵N enrichment is expressed as δ^{15} N where δ^{15} N (‰) was calculated as 1000 × (atom %_{sample} – atom %_{reference}) / atom %_{reference} and where atom %_{reference} is 0.3663%.

Groundnut plants were harvested on 14th July 1995, and divided into above - ground biomass, pods and roots. Trees were pruned on 19th May 1995 (Gliricidia trees only), and on 2nd November 1995 (all trees), and divided into leaf and stem. For harvest and pruning events the fresh weight of each sub-component was recorded, and subsamples were dried. Groundnut biomass subsamples were further divided after drying into stem and leaf, and pod subsamples were divided into husk and grain. Total dry weights were estimated for each component by multiplying total fresh weight by the subsample dry weight ratio. Each sub-component was analysed separately for ¹⁵N and % N. Plant samples were ground in increasing order of predicted ¹³N enrichment to reduce the effects of cross-contamination. Removal of ¹⁵N in plants was calculated for pruning events and at groundnut harvest. ¹⁵N removed in bi-weekly leaf samples was also quantified and included in the recovery calculation. Total ¹⁵N uptake was calculated as the sum of excess ¹⁵N removed in all plant parts. Natural ¹⁵N levels of each species were used as background levels for calculation of ¹⁵N excess. Proportional recovery of ¹⁵N was calculated as the total recovery of ¹⁵N excess in all plant parts divided by the excess ¹⁵N applied.

% N recovery

$$100 \times \left(\frac{\sum_{i}^{n} \text{Dry weight} \times \% \text{ N} / 100 \times (\delta^{-15} \text{N}_{\text{sample}} - \delta^{-15} \text{N}_{\text{background}})}{\text{N}_{\text{fertilizer}} \times (\delta^{-15} \text{N}_{\text{fertilizer}} - \delta^{-15} \text{N}_{\text{background}})}\right)$$

¹⁵N in litterfall (which was negligible), and in tree roots and stumps, was not included. No ¹⁵N enrichment was detected in groundnut samples from Zone 3 so these were not included in the analysis.

Biological nitrogen fixation

The proportions of N in groundnut and Gliricidia derived from atmospheric fixation were estimated from leaf samples taken the day before placement of ¹⁵N, using the natural abundance method (Shearer and Kohl, 1986).

% N from fixation =
$$100 \times \frac{\delta^{-15}N_{reference plant} - \delta^{-15}N_{fixing plant}}{\delta^{-15}N_{reference plant} - B}$$

Where reference plants were *Pertophorum* samples taken the day before ¹⁵N application and maize and rice samples collected in unlabelled control plots in the following season. B is the δ^{-15} N value for the fixing plant when grown on N – free medium. B values of –2.6 for groundnut shoots (G. Cadisch, 1997 unpublished) and –1.45 for *Gliricidia* prunings (Ladha et al., 1993) were used.

Statistical analysis

Analysis of variance was carried out on log or arc sin (% values) transformed values using Genstat (Payne et al., 1987) for species comparison within the same ¹⁵N application depth. Due to the limited experimental design no statistical analysis was performed on data between ¹⁵N application depths.

Results

The vertical pattern of recovery of ¹⁵N in soil samples taken 406 days after ¹⁵N application (Figure 2) showed, at least for the upper two positions, that



Figure 2. Residual $\delta^{15}N \%$ in total soil N, 406 days after placement of $({}^{15}NH_4)_2SO_4$ (20 atom $\%^{15}N$) in a hedgerow intercropping system in North Lampung, Indonesia, +/- the standard error of the means.

δ ¹⁵N %

the location of ¹⁵N was still clearly related to placement location. Differences in δ ¹⁵N between sampling depths were significant (*P* < 0.001).

All species showed a consistent increase in ¹⁵N enrichment in leaf samples taken during the first eight weeks after ¹⁵N placement at 5 cm (Figure 3). Groundnut leaves had the highest isotope enrichment at all dates after ¹⁵N placement, even from the deepest placement of 55 cm. *Gliricidia* leaves were only significantly enriched with ¹⁵N placement at 5 cm depth, whereas placement depths of 35 and 55 cm gave only a small increase in ¹⁵N above background levels. ¹⁵N enrichment of *Peltophorum* leaves increased more markedly than *Gliricidia* at all placement depths, but particularly at lower depths.

The mean total ¹⁵N recovery by 'centre' groundnut plants (0.125 m distance from the ¹⁵N application site) from placements at 5 cm depth was 13.8% of the amount of ¹⁵N applied (Table 1). Recovery declined with application depth, but even from 55 cm placements over 3% of ¹⁵N applied was recovered. In groundnut, recovery by 'border' plants (0.5 m distance from the ¹⁵N application site) was small in relation to that by 'centre' plants. In contrast, the trees recovered more similar amounts in 'border' and 'centre' zones. Recovery of ¹⁵N by *Gliricidia* trees was similar at all depths and a small percentage (< 2.1%) of that applied. *Peltophorum* trees apparently recovered most ¹⁵N from the lowest placement depth but the difference was not statistically significant due to the variability and low number of replicates.

Samples from border *Peltophorum* trees from two of the 35 cm placement depth plots showed very high enrichments, in one case resulting in calculated ¹⁵N recovery greater than the amount applied. This suggests either that the subsamples analysed for ¹⁵N content were not representative of the material from these trees, or that biomass estimates were inappropriate. Cross conta-



Figure 3. Changes in $\text{Log}_{10} \delta^{15}$ N (%c) in leaves of groundnut, *Gliricidia* and *Peltophorum* after placements of ¹⁵N at depths of 5 cm, 35 cm and 55 cm in a hedgerow intercropping system in North Lampung, Indonesia. Standard errors of the difference between means are shown.

Crop/Tree		¹⁵ N recovery (% of applied) ¹⁵ N placement depth					
		5 cm		35 cm		55 cm	SE
		Mean	Mean SE Mean SE	Mean			
Groundnut	Centre Border	13.8 1.1	3.1 0.3	8,9 1.0	3.0 0.5	3.5 0.2	0.6 0.02
Gliricidia	Centre Border	0.3 2.1	0.1 0.9	0.2	0.1	0.2 0.7	0.1 0.1
Peltophorum	Centre Border	5.0 4.5	2.2 3.8	5.6 76.7	4.5 44.6	7.8 2.0	7.8 1.7
Total		26.7		* 92.6 * 15.9		14.5	

Table 1. Measured total recovery of ¹⁵N applied (per cent) by 'centre' and 'border' plants (relative to application site) at three soil depths by groundrut and hedgerow trees *Gliricidia* and *Peltophorum* in a hedgerow intercropping system in North Lampung, Indonesia, SE = Standard error of the mean.

SE = Standard error of the mean.

* Including; ** Not including Peltophorum border recov

mination of samples during sample processing is unlikely since leaf and stem samples had similar high enrichments.

The non-fixing *Peltophorum*, maize and rice plant had higher $\delta^{15}N$ signatures than the fixing legumes and were used as reference plants for the estimation of N₂ fixation by groundnut and *Gliricidia*. *Peltophorum* may have a deeper rooting profile than the fixing legume species (Hairiah et al., 1992) and thus may obtain N from different pools. This would make it unsuitable as a reference plant if isotopic enrichment changes with soil depth (Shearer and Kohl, 1986). However, natural abundance ($\delta^{15}N$) of *Peltophorum* did not differ significantly from that in the more shallowly rooting maize and rice. Because it is not certain which is the best reference plant we expressed % N₂ fixation as the range obtained by using the different reference plants. The percentage of N in (26 day old) groundnut derived from atmospheric fixation was low (6–28%, Table 2), although this may have increased as the crop matured. *Gliricidia* derived approximately half of its N requirement from fixation.

Mean total removal of N in prunings from border and centre zones was 140 kg ha⁻¹ (*Gliricidia*) and 99 kg ha⁻¹ (*Peltophorum*) (Table 3). N production was significantly higher in *Gliricidia* (P < 0.01), but taking into account the amount of atmospheric N₂ fixation (50% or 70 kg N ha⁻¹, average of three estimates), *Gliricidia* abstracted less N from the soil. Assuming that the proportion of ¹⁵N uptake from 55 cm depth to the sum of ¹⁵N uptake from all depths was the same ratio as total soil N uptake from this depth (30% and

Table 2. Natural ¹⁵N abundance values and estimated nitrogen derived from atmospheric fixation in a hedgerow intercropping system in North Lampung. Indonesia. Standard errors are given in parentheses.

	δ ¹⁵ N %/	B*	%N derived from N ₂ fixation
Peltophorum (reference)	5.5 (0.4)		
Maize (reference)	6.0 (1.0)		_
Rice (reference)	7.9 (1.6)	-	_
Gliricidia	2.4 (0.4)	-1.45	44–58
Groundnut	5.0 (0.8)	-2.6	6-28

* B values ($\delta^{-15}N$ of plants grown in N free medium) obtained from Ladha et al. (1993) (*Gliricidia*) and Cadisch (unpublished data) (groundnut).

Table 3. Total removal of N in tree prunings, N fixation and N uptake from the soil over 203 days in a hedgerow intercropping system in North Lampung, Indonesia. Standard errors are given in parentheses.

	N in prunings	N fixation	N uptake from soil [#]	N uptake from safety-net zone"	
	$(kg ha^{-1})$	(kg ha ⁻¹)	(kg ha⁻¹)	(kg ha ⁻¹)	
Gliricidia	140 (8.9)	70	70	21	
Peltophorum	99 (6.7)	0	99	42	
SED	11.2**		8.1**		

^{*} Total uptake of soil N was calculated assuming that *Gliricidia* obtains 50% of its N from fixation. Uptake from the safety-net zone was deduced from the proportion of ¹⁵N recovery below 55 cm depth.

42% for *Gliricidia* and *Peltophorum* respectively), total soil N uptake from this safety-net zone (55 cm) was 21 kg ha⁻¹ for *Gliricidia* and 40 kg ha⁻¹ for *Peltophorum*.

Discussion

Atkinson et al. (1978) used ¹⁵N placement in orchard tree based systems previously but their aim was to evaluate fertilizer N use efficiency (herbicide vs. grass cover) and the deepest application was 0.2 m. Our results suggest that direct ¹⁵N placement has great potential for study of root activity in agroforestry systems as well as giving indications about the potential role of tree roots to act as a safety-net. The method of applying ¹⁵N with a carbon source was successful in restricting its movement within the soil (Figure 2), which is essential to attribute root activity to defined rooting depths. ¹⁵N enrichments obtained were sufficiently above background natural enrichments to quantify percentage recovery, particularly considering the ability of modern mass spectrometers to detect small differences in isotope enrichment. While treatment variation was acceptable in the groundnut crop, variation was large with the trees. This is likely to have resulted in part from heterogeneity in root distributions, and highlights the need for larger numbers of replicates for each treatment for future applications. This heterogeneity may be more pronounced in the centre of the crop alley, and ¹⁵N application closer to the hedgerow may give higher and more consistent values for safety-net recovery.

The data set presented here is consistent with the hypothesis that *Peltophorum* takes up more N from deeper soil layers than *Gliricidia*. While ¹⁵N recovery by groundnut decreased sharply with increasing depth of ¹⁵N placement, recovery of ¹⁵N by 'centre' trees (opposite ¹⁵N placements) was similar at all depths. *Peltophorum* roots maintained their high activity at all depths and apparently took up more ¹⁵N than *Gliricidia*. However, the high enrichment of some 'border' *Peltophorum* trees highlights the heterogeneity in root distributions. *Peltophorum* roots may extend 4 m or more from the stem (Rowe, pers. obs.) and individual trees may show a considerable variation in ¹⁵N recovery according to the location of their roots. This heterogeneity in root distribution is likely to increase with distance from the tree, thus reduced variability in ¹⁵N recovery may be obtained by applying the isotope closer to the tree base. Measuring recovery of ¹⁵N by individual trees rather than as a mean of several trees may give a clearer indication of the heterogeneity of root systems.

The second hypothesis, that *Peltophorum* competes less strongly than Gliricidia for topsoil N, is not supported by the data. Gliricidia recovered very little ¹⁵N from any placement depth. This indicates that it did not compete strongly with groundnut for N in this case. However, since N recovery by trees was calculated from a pruning nearly four months after groundnut harvest, the conclusions that can be drawn about competition within the growth phase of the crop are limited. The small ¹⁵N recovery by *Gliricidia* suggests that it derived a high proportion of nitrogen from atmospheric fixation, and this is confirmed by the natural ¹⁵N abundance estimate of 44–58% N₂ fixation (Table 2). Gliricidia apparently has taken up a smaller proportion of its soil derived N from labelled pools than Peltophorum, perhaps because it has a smaller proportion of roots in the centre of the alley where ¹⁵N was placed (Table 3). To take into account variation in root activity with distance from the tree it would be best to inject ¹⁵N at points across the alley to obtain an integrated N uptake activity measurement. The similar root distributions of crops and *Gliricidia* found on this site by Akiefnawati (1995), with proportionally high topsoil and low subsoil root length densities, suggests a niche overlap and consequent strong competition for N (Vandermeer, 1990). Schroth and Zech (1995) argued that Gliricidia tree root length densities were too low to cause strong competition with the crop for soil resources but their measured root length density was lower than those found in our study area. Total N uptake by *Gliricidia* in this system was substantial and would represent considerable competition if *Gliricidia* were not obtaining N from another source. Overlap of uptake distribution for other nutrients and for water is likely, however, and *Gliricidia* may be expected to compete strongly for these resources.

Uptake of ¹⁵N by groundnut declined with the depth of ¹⁵N placement, but 3% of applied ¹⁵N was still recovered from 55 cm placement depth, showing that groundnut roots do penetrate the subsoil. This may be because decaying tree roots in this established system have provided channels where aluminium toxicity is less extreme (van Noordwijk et al., 1991). There is however substantially less uptake from this depth by 'border' plants, suggesting that deep roots extend horizontally less than shallow roots.

Conclusions

The study provides insights into the distribution of root activity in a hedgerow intercropping system, and its influence on the degree of complementarity in the system. The evidence from ¹⁵N in soil after 406 days suggests that the placement method used successfully maintained labelled N within the target soil layer. Enrichment of plant materials can therefore be traced with some degree of certainty to uptake from this soil layer. The potential for *Peltophorum* to act as a safety – net has been shown, since it is actively taking up a substantial proportion (42%) of its N from deeper soil layers. The results also suggest that root distribution is not the sole factor explaining N uptake distribution. For example, no evidence was found that *Gliricidia* was in strong competition with the crop for topsoil N, despite its predominantly shallow root distribution. Indeed, *Gliricidia* recovered little ¹⁵N from any depth. This is in part because of the large proportion of N obtained from atmospheric fixation, but *Gliricidia* must also have taken up a smaller proportion of its soil-derived N from labelled pools.

A more detailed measurement of ¹⁵N uptake activity across the alley would be needed to calculate a reliable balance for N and assess the degree of competition with the crop for N by these two trees in the whole system. Management techniques, such as the additional pruning which was applied to *Gliricidia* trees, are likely to have a major effect on the root activity pattern in space and time. However, the method used will assist with developing a mechanistic understanding of nitrogen dynamics in mixed cropping systems, and combined with WaNuLCAS, allows the safety – net efficiency of contrasting trees to be evaluated (Cadisch et al., 1997).

Acknowledgements

This publication is an output from a research project funded by the Department for International Development of the United Kingdom. However, the Department for International Development can accept no responsibility for any information provided or views expressed (R6523, Forestry Research Programme).

References

- Akiefnawati R (1995). Pengaruh naugan, kompetisi serapan air dan hara tanaman pagar terhadap pertumbuhan dan produksi jagung pada ultisol daerah Lampung Utara. MSc Thesis, Universitas Brawijaya, Malang
- Atkinson D, Johnson MG. Mattam D and Mercer ER (1978) The effect of orchard soil management on the uptake of nitrogen by established apple trees. J Sci Food Agric. 30: 129-135
- Cadisch G, Rowe EC and van Noordwijk M (1997) Nutrient harvesting the tree root safety net. Agrofor Forum 8: 31-33
- Cannell MGR, van Noordwijk M and Ong CK (1996) The central agroforestry hypothesis: the trees must acquire resources that the crop would not otherwise acquire. Agrofor Syst 34: 27-31
- Hairiah K, van Noordwijk M. Santoso B and Syekhfani MS (1992) Biomass production and root distribution of eight trees and their potential for hedgerow intercropping on an ultisol in southern Sumatra. Agrivita 15: 54-68
- Hairiah K, Van Noordwijk M and Setijono S (1991) Tolerance to acid soil-conditions of the velvet beans Mucuna pruriens var utilis and M. deeringiana. 1. Root development. Plant Soil 134: 95-105
- Handayanto E, Giller KE and Cadisch G (1997) Regulating N release from legume tree prunings by mixing residues of different quality. Soil Biology and Biochemistry 29: 1417–1426
- IAEA (1975) Root activity patterns of some tree crops. International Atomic Energy Commission, Vienna, Austria
- Ladha JK, Peoples MB, Garrity DP, Capuno VT and Dart PJ (1993) Estimating dinitrogen fixation of hedgerow vegetation using the nitrogen-15 natural abundance method. Soil Sci Soc Am J 57: 732-737
- Payne RW, Lane PW, Ainsley AE, Bicknell KE, Digby PGN, Harding SA, Leech PK, Simpson HR, Todd AD, Verrier PJ and White RP (1987) Genstat 5 reference manual. Clarendon Press, Oxford, UK
- Schroth G and Zech W (1995) Root length dynamics in agroforestry with Gliricidia sepium as compared to sole cropping in the semi-deciduous rainforest zone of West Africa. Plant Soil 170: 297-306
- Shearer G and Kohl DH (1986) N₂ fixation in field settings: estimates based on natural abundance. Australian J of Plant Physiol 13: 699–756
- Van der Heide J, Setijono S, Syekhfani MS, Flach EN, Hairiah K, Ismunandar S, Sitompul SM and Van Noordwijk M (1992) Can low external input cropping systems on acid upland soils in the humid tropics be sustainable? Backgrounds of the UniBraw/Nitrogen management project in Bunga Mayang (Sunkai Selatan, Kotabumi, S. Sumatera, Indonesia). Agrivita 15: 1-10
- van Noordwijk M (1996) Models as part of agroforestry research design. Agrivita 19: 192-197
- van Noordwijk M, Hairiah K, Lusiana B and Cadisch G (1997a) Tree-Soil-Crop Interactions in Sequential and Simultaneous Agroforestry Systems. In: Bergstrom L and Kirschner H (eds) Carbon and Nutrient Dynamics in Natural and Agricultural Tropical Ecosystems, pp 173-190. CAB International, Wallingford, UK
- van Noordwijk M, Hairiah K. Partoharjono S, Labios RV and Garrity DP (1997b) Food-cropbased production systems as sustainable alternatives for Imperata grasslands? Agrofor Syst 36: 55-82
- van Noordwijk M, Lawson G. Groot JJR and Hairiah K (1996) Root distribution in relation to nutrients and competition. In: Ong CK and Huxley PA (eds) Tree-Crop Interactions - a Physiological Approach. pp 319-364. CAB International.
- van Noordwijk M and Lusiana B (1997) WaNuLCAS, a model of water. light and nutrient capture in agroforestry systems. Agroforestry Systems (this issue)
- van Noordwijk M, Sitompul SM, Hairiah K and Ismunandar S (1992) Nitrogen management under high rainfall conditions for shallow rooted crops: principles and hypotheses. Agrivita 15: 11-19

- van Noordwijk M. Widianto, Heinen M and Hairiah K (1991) Old tree root channels in acid soils in the humid tropics; important for crop root penetration, water infiltration and nitrogen management. Plant Soil 134: 37-44
- Vandermeer J (1990) The ecology of intercropping. Cambridge University Press, Cambridge, UK
- Witty JF and Ritz K (1984) Slow-release ¹⁵N fertilizer formulations to measure N₂-fixation by isotope dilution. Soil Biol and Biochem 16: 657–661
- Wong MTF, Hughes R and Rowell DL (1990) Retarded leaching of nitrate in acid soils from the tropics: measurement of the effective anion exchange capacity. J Soil Sci 41: 655-663
 Xu ZH, Saffinga PG, Myers RJK and Chapman AL (1993) Nitrogen cycling in leucaena
- (Leucaena leucocephala) alley cropping in semi-arid tropics. Plant Soil 148: 63–72
- Young A (1997) Agroforesty for Soil Management. 2nd Edn. CAB International, Wallingford, UK